



Chemical profile of the parotoid gland secretion of the Amazonian toad (*Rhinella margaritifera*)

Adilson Paulo Senhorin^{b,*}, Jacqueline Kerkhoff^{a,b}, Evandro Luiz Dall'Oglio^{a,c}, Domingos de Jesus Rodrigues^b, Leonardo Gomes de Vasconcelos^c, Valéria Dornelles Gindri Senhorin^b

^a Programa de Pós-Graduação Rede de Biodiversidade e Biotecnologia da Amazônia Legal - PPG-BIONORTE, Coordenação Geral do Doutorado em Biodiversidade e Biotecnologia, Universidade Estadual do Maranhão, Cidade Universitária Paulo VI, Predio da Veterinária. Av. Lourenço Vieira da Silva, nº 1000, CEP: 65.055-310, São Luís, MA, Brazil

^b Laboratórios Integrados de Pesquisa em Química (LIPEQ), Programa de Pós-Graduação em Ciências Ambientais, Instituto de Ciências Naturais, Humanas e Sociais, Universidade Federal de Mato Grosso, Campus de Sinop. Avenida Alexandre Ferronato, nº 1200, Bairro Setor Industrial, CEP 78557-267, Sinop, Mato Grosso, Brazil

^c Departamento de Química, Instituto de Ciência Exatas e da Terra, Universidade Federal de Mato Grosso – UFMT, Av. Fernando Corrêa da Costa, nº 2367, Bairro Boa Esperança, Cuiabá, MT, 78060-900, Brazil

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ABSTRACT

The secreted poisonin bufonids (Anura: Bufonidae) include proteins, biogenic amines, toxic bufadienolides and alkaloids. The chemical composition of the methanolic extract of parotoid gland secretions by the Amazonian toad *Rhinella margaritifera* was evaluated in a UFLC-DAD-micrOTOF system. Of the twenty three compounds found in the methanolic extract, eighteen were identified by the mass/charge ratio as: five arginine diacids, six bufagenins (telocinobufagin, marinobufagin, bufotalin, cinobufotalin, bufalin and cinobufagin), six bufotoxins, and an alkaloid (dehydrobufotenin).

The Bufonidae family, which includes the so-called true toads, is composed of 52 genera with worldwide distribution (AmphibiaWeb, 2020). These animals occur in a wide variety of habitats from deserts to tropical forests.

Bufoiids produce very potent toxins in their skin, especially concentrated in the parotoid glands, dorsal structures located in the post-orbital region, and can be fatal to predators when ingested (Clarke, 1997; Toledo et al., 1992). However, several studies have been exploring these compounds in pharmacological models for antitumor and cytotoxic, cardiotoxic, antifungal, antimicrobial and antiparasitic activity (Barnhart et al., 2017; Chen et al., 2018; Cunha-Filho et al., 2010; Medeiros et al., 2019; Ferreira et al., 2013; Banfi et al., 2016; He et al., 2019; Kowalski et al., 2018; Meng et al., 2019; Nalbantsoy et al., 2016; Perera-Córdova et al., 2016; Rodríguez et al., 2017; Sousa et al., 2017).

The *Rhinella margaritifera* Laurenti (*R. margaritifera*) (Laurenti, 1768) species has wide distribution throughout the Amazon basin, which includes Colombia, Venezuela, Peru, Bolivia, Brazil and the Guyana. It is a terrestrial species with daytime and nighttime activity, which lives in

the litterfall of primary forest (Ávila et al., 2010). It is a brown toad (Fig. 1) of medium size (males measure between 40 and 67 mm and females between 46 and 76 mm), with conspicuous parotoid glands and developed cranial ridges (especially in females). The morphology and brown coloration of *R. margaritifera* allows them to mix with the litterfall. Individuals climb to low vegetation up to 1.5 m above the ground at night, where they remain inactive (Moravec et al., 2014). Adults feed on arthropods, especially ants (Lima and Magnusson, 2000).

Parotoid glands from toads can secrete several different compounds, including proteins, biogenic amines, alkaloids and steroids (Clarke, 1997; Daly et al., 2005; Mariano et al., 2018; Rash et al., 2011; Zhao et al., 2006). The components abundance and diversity of each of these compounds class can vary according to the life history of the amphibian in question, gender, and season (Sherman et al., 2009).

Bufoiadienolides are the major components of bufonid parotoid secretions. These molecules are polyhydroxy-steroids with 24 carbon atoms, characterized by an unsaturated lactone ring and an α -pyrone group linked to C-17. They can be found in free form or in conjunction

* Corresponding author.

E-mail addresses: sinhorin.adilson@gmail.com, sinhorin@ufmt.br (A.P. Senhorin), jackerkhoff@gmail.com (J. Kerkhoff), dalloglio.evandro@gmail.com (E.L. Dall'Oglio), djmingo23@gmail.com (D. de Jesus Rodrigues), vasconceloslg@gmail.com, vasconceloslg@ufmt.br (L.G. de Vasconcelos), valeriadgindri@gmail.com (V.D.G. Senhorin).

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Fig. 1. *R. margaritifera*. Photography by domingos rodrigues.

with sulfates, dicarboxylic esters and amino acids at the C-3 position (Sousa et al., 2017). These possibilities lend this group of compounds to have enormous structural diversity (Gao et al., 2010).

These steroids are potent inhibitors of Na^+/K^+ -ATPase activity, and if ingested they cause a bitter taste in the mouth, nausea or heart failure. In addition, they repel and may even kill predators. Bufadienolides can also contribute to the toads' immunological defense against pathogens (Barnhart et al., 2017).

Although several trivial nomenclatures for bufadienolides can be found in the literature, we have adopted the nomenclature of the bufagenins and bufotoxins subclass for the purposes of this work. Thus, the smaller hydrolyzed bufadienolide molecules are called bufagenins (free bufadienolides), and the larger bufadienolide molecules which have a side chain of amino acids are called bufotoxins.

About 100 bufadienolides have been reported in the literature, being bufonid poisons (Steyn and Van Heerden, 1998). Although amphibian skin secretions have proven to be a rich source of unique molecules, they remain largely unexplored and represent great potential for developing new molecular models for pharmacological and toxicological evaluations and even for synthesis and medicinal chemistry. Therefore, the objective of this work was to analyze the chemical composition of *R. margaritifera* and to trace the bufagenin and bufotoxin profile by chromatographic and spectrometric analyzes for the first time.

The *R. margaritifera* specimen was captured in January 2018 in the municipality of Cotriguaçu (9°49'20.00"S 58°17'16.00" W) in Mato Grosso state, Brazil, identified by the team of biologists from the Federal University of Mato Grosso, under the coordination of Prof. Dr. Domingos de Jesus Rodrigues (IBAMA, SISBIO: 30034-1). The poison was collected by manually compressing the animal's parotoid glands and then stored in glass flasks containing silica gel. The toad was returned to nature after collecting the poison. The access to Brazilian Genetic Heritage was registered with SisGen, in compliance with the provisions of Law No. 13,123/2015 and its regulations, under No. ACC9622. The dried poison was crushed with a mortar and pestle and extracted in an ultrasonic bath with 1:10 methanol for 60 min (Kerkhoff et al., 2016). The obtained methanolic extract was dried by rotary evaporation at reduced pressure of 600 mmHg at 40 °C and finished in a high vacuum pump. The obtained extract was subsequently stored at 4 °C. For chromatographic analysis, 1 mg of methanolic extract was solubilized in 1 mL of HPLC grade methanol and filtered through a filter membrane with porosity of 0.22 μm.

The analyzes were performed in a Liquid Chromatography System with Detection by Ultraviolet Spectroscopy and Mass Spectrometry (UFLC-DAD-micrOTOF) and the method was adapted from the one developed by Schmeda-Hirschmann et al. (2016) and had the following

conditions: Injection Volume: 1 μL; Chromatographic column: Kromasil 100-5-C18 column, 250 × 4.6 mm, 5 μm; Pre-Column: Kromasil Guard Column 100-5-C18, 4.6 × 10 mm, 5 μm; Chromatographic Conditions: Mobile Phase (Solvent A: 0.5% Formic Acid Solution; Solvent B: Acetonitrile 0.5% formic acid); Elution Mode: Gradient (following the following schedule: 0–45 min with 8–64% Solvent B); Flow: 1.0 mL/min; Column Temperature: 40 °C.

Chromatographic analysis of the methanolic extract obtained from the *R. margaritifera* parotoid secretion detected 23 compounds (Fig. 2), in which 18 of these were identified by the mass/charge ratio (Table 1). Compounds 1–9 are smaller, polar molecules which correspond to the amino acid arginine and arginine diacids, in addition to an alkaloid, dehydrobufotenin. The compounds 10–23 are those derived from cardiotonic steroids, of bufagenin and bufotoxin types, with less polarity.

Compound 1 was identified as the amino acid Arginine, with the molecular formula of $\text{C}_6\text{H}_{14}\text{N}_4\text{O}_2$ and m/z 175.1187 (Shek et al., 2006). Compounds 2, 3, 5 and 6 have been identified as arginine diacids. The compounds belong to a homologous series which varies in the size of the diacid carbon chain, between 6 and 9, and have been designated as adipoyl-, pimeloyl-, suberoyl- and azelalyl-argininyl. This identification proposal is in accordance with the results of the study by Schmeda-Hirschmann (2014).

Compound 4 was identified as the Dehydrobufotenin alkaloid ($\text{C}_{12}\text{H}_{14}\text{N}_2\text{O}$; m/z 203.1169), a tricyclic metabolite of serotonin and has been identified in several species of the Bufonidae family (Schmeda-Hirschmann et al., 2017).

The bufagenins identified in this work correspond to the compounds 10, 15, 16, 18, 20 and 23, and are described as follows: compound 10 was identified as Telocinobufagin ($\text{C}_{24}\text{H}_{34}\text{O}_5$; m/z 403.2449); compound 15 corresponds to Marinobufagin ($\text{C}_{24}\text{H}_{32}\text{O}_5$; m/z 401.2319); compound 16 was identified as Bufotalin ($\text{C}_{26}\text{H}_{36}\text{O}_6$; m/z 445.2578); compound 18 was identified as Cinobufotalin ($\text{C}_{26}\text{H}_{34}\text{O}_7$; m/z 459.2381); compound 20 was assimilated to Bufalin ($\text{C}_{24}\text{H}_{34}\text{O}_4$; 387.2535); and compound 23 was identified as Cinobufagin ($\text{C}_{26}\text{H}_{34}\text{O}_6$; m/z 443.2419) (He et al., 2019). All ions and fragments are consistent with those found in the work of Zhang et al. (2016), who performed chromatographic analyzes of 64 bufadienolides found in bufonids. These molecules have already been described in several studies which investigated the chemical composition of bufonid poisons, such as *Rhinella marina*, *Rhinella schneideri*, *Rhinella ornata*, *Rhinella arenarum*, *Bufo garzarizans*, and *Peltophryne fustiger* (Chen et al., 2018; Cunha-Filho et al., 2010; Perera-Córdova et al., 2016; Petroselli et al., 2018; Schmeda-Hirschmann et al., 2017, 2016).

Bufotoxins are bufadienolides with an amino acid side chain. Thus, there are numerous possible combinations of bufadienolides, diacids and amino acids. A fragmentation pattern was found for the bufotoxins, with the most intense fragment being arginine diacid, followed by the loss of one amine group and two H_2O groups of that fraction, generating the fragments which correspond to $[\text{M} + \text{H} - \text{NH}_2]$, $[\text{M} + \text{H} - \text{NH}_2 - \text{H}_2\text{O}]$ and $[\text{M} + \text{H} - \text{NH}_2 - 2\text{H}_2\text{O}]$, respectively. Thus, compound 11 was identified as 3-(*N*-adipoyl argininyl) bufotalin ($\text{C}_{38}\text{H}_{56}\text{N}_4\text{O}_{10}$); compound 12 was also identified as a derivative of bufotalin, 3-(*N*-pimeloyl argininyl) bufotalin ($\text{C}_{39}\text{H}_{58}\text{N}_4\text{O}_{10}$; m/z 743.4168); compound 13 was identified as 3-(*N*-suberoyl argininyl) telocinobufagin ($\text{C}_{38}\text{H}_{60}\text{N}_4\text{O}_{10}$; m/z 715.4246); 17 was assimilated to 3-(*N*-suberoyl argininyl) bufotalin ($\text{C}_{40}\text{H}_{60}\text{N}_4\text{O}_{10}$; m/z 757.4392); compound 19 was identified as 3-(*N*-pimeloyl argininyl) cinobufagin ($\text{C}_{39}\text{H}_{56}\text{N}_4\text{O}_{10}$; m/z 741.4064); and compound 21 corresponds to as 3-(*N*-suberoyl argininyl) cinobufagin ($\text{C}_{40}\text{H}_{58}\text{N}_4\text{O}_{10}$; m/z 755.4231) (Schmeda-Hirschmann et al., 2017).

The major compounds were the bufadienolides Bufotalin (Compound 16, 33%), Cinobufagin (Compound 23, 16%) and Marinobufagin (Compound 15, 13%), which correspond to more than 60% of the extract. These findings corroborate the high toxicity found in bufonid poisons (Tóth et al., 2019). Altogether, 80% of the constituents of *R. margaritifera* extract were identified. It was not possible to find data in

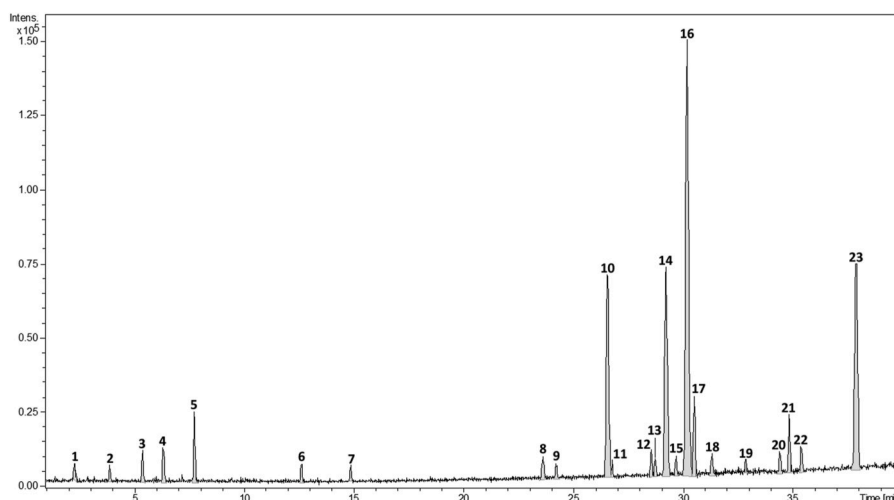


Fig. 2. UFLC-DAD-microTOF chromatogram of *R. margaritifera* methanolic extract. Chromatographic Conditions: Mobile Phase (Solvent A: 0.5% Formic Acid Solution; Solvent B: Acetonitrile 0.5% formic acid); Elution Mode: Gradient (following the schedule: 0–45 min with 8–64% Solvent B; Flow: 1.0 mL/min; Column Temperature: 40 °C.

Table 1

Identification of the methanolic extract constituents of *R. margaritifera* poison by UFLC-DAD-microTOF.

Compound	Rt (min)	Relative %	[M+H] ⁺	Fragmentation	Identification
1	2.4	0.8	175.1187	130.1568; 116.0473; 112.0278; 105.0385	Arginine ^g
2	4.0	0.6	303.1652	226.0988; 175.1182; 158.0920; 116.0720	Adipyl arginine ^{d,f}
3	5.4	1.3	317.1817	175.1173; 158.0920; 116.0728; 112.0728	Pimeloyl arginine ^{d,f}
4	6.4	1.7	203.1169	188.0945; 173.0705; 160.0986; 145.0753	Dehydrobufotenin ^{a,b,d,f}
5	7.8	2.9	331.1972	175.1148; 158.0926; 130.0970; 116.0707	Suberoyl arginine ^{b,d,f}
6	12.7	0.8	345.2134	175.1201; 158.0924; 116.0718; 112.0884	Azelayl arginine ^{d,f, e}
7	14.9	0.5	197.0778	175.0965; 162.0548; 157.0860; 112.0299	Not assigned
8	23.6	1.3	211.0916	189.1108; 173.0792; 171.0973; 169.0601	Not assigned
9	24.2	0.8	467.2059	369.2434; 351.2286; 255.2089; 147.1110	Not assigned
10	26.6	13.1	403.2449	367.2253; 215.1759; 161.1296; 151.0378	Telocinobufagin ^{a,b,d,f}
11	26.8	0.5	729.4028	303.1642; 286.1379; 268.1243; 250.1150	3-(N-adipoyl argininy)l bufotalin ^g
12	28.6	1.0	743.4168	317.1792; 300.1537; 282.1406; 264.1311	3-(N-pimeloyl argininy)l bufotalin ^g
13	28.7	0.6	715.4246	331.1967; 314.1674; 278.1532; 175.1183	3-(N-suberoyl argininy)l telocinobufagin ^{b,d}
14	29.2	15.0	431.2413	349.2149; 187.0742; 175.1470; 161.1305	Not assigned
15	29.7	0.8	401.2319	365.2129; 215.1770; 157.0955; 149.1283	Marinobufagin ^{a,b,d,f}
16	30.2	32.8	445.2578	367.2244; 349.2155; 241.1212; 161.1309	Bufotalin ^{g, e}
17	30.5	3.6	757.4392	331.1967; 296.1623; 278.1494; 175.1199	3-(N-suberoyl argininy)l bufotalin ^g
18	31.3	1.1	459.2381	381.2106; 363.1969; 255.1000; 201.1627	Cinobufotalin ^c
19	32.8	0.4	741.4064	317.1827; 282.1473; 264.1333; 175.1188	3-(N-pimeloyl argininy)l cinobufagin ^f
20	34.4	1.0	387.2535	369.2392; 351.2325; 255.2078; 147.1169	Bufalin ^{a,b,d,f}
21	34.8	2.4	755.4231	331.1986; 314.1724; 278.1512; 175.1198	3-(N-suberoyl argininy)l cinobufagina ^f
22	35.4	1.2	274.2734	274.2702; 256.2652; 106.0889; 102.0928	Not assigned
23	37.9	15.8	443.2419	365.2082; 215.1784; 187.1476; 151.0390	Cinobufagin ^{a,h}

^a Ferreira et al., (2013).

^b Schmeda-Hirschmann et al., (2014).

^c Zhang et al., (2016).

^d Schmeda-Hirschmann et al., (2016).

^e Schmeda-Hirschmann et al., (2017).

^f Schmeda-Hirschmann et al., (2017).

^g Petroselli et al., (2018).

^h He et al., (2019).

^e Perera-Córdova et al., (2016).

the literature for identifying the compounds 7, 8, 9, 14 and 22, and their fragments.

The chemical analysis of the methanolic extract of the secretions of the parotid glands of *R. margaritifera* resulted in the profile of bufagenins and bufotoxins that composes this poison, with Bufotalin as the major compound. The results point to the relevance of new investigations in toads of the *Rhinella* genus and to establish a reference for other poisons of Amazonian bufonids.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

CRediT authorship contribution statement

Adilson Paulo Sinhorin: Project administration, Writing - review & editing. **Jacqueline Kerkhoff:** Methodology, Validation. **Evandro Luiz Dall'Oglio:** Supervision. **Domingos de Jesus Rodrigues:** Funding

acquisition. **Leonardo Gomes de Vasconcelos:** Data curation. **Valéria Dornelles Gindri Sinhorin:** Writing - review & editing.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.toxicol.2020.04.106>.

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