



**UNIVERSIDADE FEDERAL DO AMAZONAS**  
**INSTITUTO NACIONAL DE PESQUISAS DA AMAZÔNIA**  
**PROGRAMA DE PÓS-GRADUAÇÃO EM ZOOLOGIA**

**Efeitos do ambiente sobre estimativas de diversidade taxonômica, funcional e filogenética de anuros em bancos de macrófitas lacustres na Amazônia Central**

**Discente: Mahima Hemnani**

**Manaus, Amazonas**

**Novembro/2021**



**UNIVERSIDADE FEDERAL DO AMAZONAS**  
**INSTITUTO NACIONAL DE PESQUISAS DA AMAZÔNIA**  
**PROGRAMA DE PÓS-GRADUAÇÃO EM ZOOLOGIA**

**Efeitos do ambiente sobre estimativas de diversidade taxonômica, funcional e filogenética de anuros em bancos de macrófitas lacustres na Amazônia Central**

**Discente: Mahima Hemnani**

**Orientador: Prof. Dr. Igor Luis Kaefer**

**Coorientador: Prof. Dr. Rafael de Fraga**

Dissertação de Mestrado apresentada à Universidade Federal do Amazonas como parte dos requisitos para obtenção do título de Mestre pelo Programa de Pós-Graduação em Zoologia

**Manaus, Amazonas**  
**Novembro/2021**

## Ficha Catalográfica

Ficha catalográfica elaborada automaticamente de acordo com os dados fornecidos pelo(a) autor(a).

H489e Hemnani, Mahima  
Efeitos do ambiente sobre estimativas de diversidade taxonômica, funcional e filogenética de anuros em bancos de macrófitas lacustres na Amazônia Central / Mahima Hemnani . 2021  
49 f.: il. color; 31 cm.

Orientador: Igor Luis Kaefer  
Coorientador: Rafael de Fraga  
Dissertação (Mestrado em Zoologia) - Universidade Federal do Amazonas.

1. Amphibia. 2. Ecologia de comunidades. 3. Lagos amazônicos. 4. Processos de nicho. 5. Processos neutros. I. Kaefer, Igor Luis. II. Universidade Federal do Amazonas III. Título

## **AGRADECIMENTOS**

Início esta seção agradecendo ao meu eterno orientador Prof. Dr. Marcelo Menin (*in memoriam*), vitimado pela pandemia da COVID-19, que foi e sempre será amado por todos nós! Agradeço por todos os ensinamentos durante a graduação e sou imensamente grata por sua orientação durante o planejamento do projeto e a coleta de dados do mestrado. Levarei as lembranças dele para o resto da minha vida, onde quer que eu esteja.

Ao meu orientador, Prof. Dr. Igor Kaefer: agradeço imensamente por todo o apoio que sempre me proporcionou, pela orientação, pela compreensão, pela liberdade intelectual, pelo encorajamento, pelas piadas, pelas conversas aleatórias e por ter acreditado no meu trabalho desde a primeira vez que entrei no laboratório ainda no início da graduação. Apesar de todos os percalços ocorridos durante o mestrado, ele sempre tentou me ajudar da melhor forma possível, não só procurando soluções, mas principalmente nunca desistindo de mim! Muito obrigada por tudo.

Agradeço ao meu coorientador Dr. Rafael de Fraga (Rato), que aceitou fazer parte da orientação no meio do projeto e sob circunstâncias tão inesperadas. Muitíssimo obrigada por tudo, pelo suporte e colaboração em muitos pontos do desenvolvimento deste trabalho e pelo feedback constante ao longo da pesquisa.

Aos meus orientadores, para sempre, toda a minha gratidão por investir tempo na minha formação científica e humanística. Aos coordenadores do PPG Zoologia Prof. Dr. Marcelo Menin (*in memoriam*), Prof. Dr. Fabricio Baccaro e Profa. Dra. Cintia Cornelius por todo o apoio ao longo desta jornada.

Ao Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) pelo financiamento das atividades de pesquisa por meio do Projeto Universal "Anfíbios da Amazônia Brasileira: taxonomia, distribuição e ecologia" (Processo # 405640/2016-1), à Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) e à Fundação de Amparo à Pesquisa do Estado do Amazonas (FAPEAM) pelo apoio financeiro ao PPG Zoologia, e à FAPEAM pela bolsa de estudos concedida a mim.

Agradeço aos meus pais, Jasmine e Dev, por sempre me apoiarem nos meus sonhos. Se não fosse por acreditarem que a educação é a melhor herança que pode ser deixada para um filho, eu não estaria aqui.

À minha querida irmã mais nova, Jogan, que toda vez tem que me aguentar quando estou querendo “surtar” e com isso acabar brigando com todo mundo. Ela é única, pois me atura de uma forma inimaginável. Ela é minha melhor amiga e sem ela eu não teria mais sanidade mental (ou um trabalho constantemente corrigido).

Aos meus amigos mais próximos, Rubia, Iza, Raielle, Pedro e Francisco, agradeço o amor, companheirismo, colaboração, risadas aleatórias, por me acompanharem nos trabalhos em campo, e especialmente quando tudo parecia estar dando errado. As conversas com vocês deixam meu dia mais leve e o apoio constante de vocês me faz sentir que tudo é possível nessa vida.

Aos professores da Universidade Federal do Amazonas, em especial aos do Instituto de Ciências Biológicas, toda a minha admiração. São vocês que inspiram as novas gerações a traçar os tortuosos e fascinantes caminhos da pesquisa, ensino e extensão. Muitos dos professores excepcionais que tive o prazer de conhecer moldaram definitivamente a forma como eu vejo o mundo e a ciência.

Por fim, aos meus avós, por me guiarem e me apoiarem mesmo estando distantes. Em especial, ao meu avô. Saudades de você. Sei que está cuidando de mim onde quer que esteja.

Agradeço a todas as forças, divinas ou não, que possibilitaram a conclusão com êxito desse trabalho, e espero que ele possa abrir novos caminhos tanto para mim quanto para esta fantástica área do conhecimento que é a ecologia de comunidades. Muito obrigada a todos que colaboraram com essa jornada científica. Suas contribuições nunca serão esquecidas.

## LISTA DE FIGURAS

Figura 1: Study area in the Catalão lake, located in Central Amazonia, Brazil. Yellow circles are 50 x 5 m sampling units.....	18
Figura 2: Phylogenetic tree showing the genetic relationship between anurans found at the Catalão Lake (adapted from Jetz & Pyron 2018) .....	24
Figura 3: Partials derived from multiple linear models tested to quantify the effects of environmental variables on frog $\alpha$ - diversity (A, B, C), $\beta$ -diversity (D, E, F), functional diversity (G, H, I) and phylogenetic diversity (J, K, L) estimates. The more informative environmental variables for each metric (based on the stepwise regression) are in the image.....	30
Figura 4: Fourth corner interaction between amphibian species traits and environmental variables. Positive and negative associations are shown respectively in blue and red.....	31

## **LISTA DE TABELAS**

Tabela 1: List of functional traits and their ecological meanings. CO = continuous, CA = categorical, OB = observed, LT = literature.....	21
Tabela 2: Minimum (Min), maximum (Max), mean and standard deviation (SD) values of environmental variables measured in 50 plots distributed along the Catalão Lake, Central Amazonia.....	26
Tabela 3: Frog species found in the 50 plots distributed at the Catalão Lake, Central Amazonia.....	28

## RESUMO

Assembleias podem ser estruturadas por processos neutros, baseados em nicho, ou por combinações entre eles. Compreender a influência desses processos e avaliar medidas de diversidade como métricas taxonômicas, funcionais e filogenéticas na estrutura de assembleias ecológicas pode mostrar padrões e desvendar mecanismos responsáveis por suas montagens. Os anuros (pererecas, rãs e sapos) representam uma grande parte da diversidade de vertebrados em ambientes tropicais e são um grupo-chave na avaliação de padrões de diversidade em diferentes escalas espaciais. Eles podem ser terrestres, aquáticos e encontrados em bancos de macrófitas. Estes bancos consistem em emaranhados de plantas aquáticas e semiaquáticas enraizadas ou livremente flutuantes que fornecem abrigo, alimentação, bem como locais para reprodução, desova e desenvolvimento para muitos organismos, incluindo anuros, e podem promover dispersão e fluxo gênico da fauna por longas distâncias. Dado que bancos de macrófitas podem ter influência na estrutura da diversidade de anuros, nosso estudo teve como objetivo testar a influência de fatores espaciais e ambientais na diversidade taxonômica, funcional e filogenética de assembleias de anuros encontradas nestes locais. Nosso estudo foi realizado no Lago do Catalão, Amazônia Central. Amostramos 50 parcelas de 50 por 5 m durante o período de enchente, em margens de bancos de macrófitas distribuídas ao longo do lago. Variáveis geográficas e ambientais foram mensuradas em seis pontos equidistantes de cada parcela. Mostramos que a altura das macrófitas influencia a diversidade taxonômica  $\alpha$ ; a altura das macrófitas e a distância da margem mais próxima influenciam a diversidade taxonômica  $\beta$ ; o pH da água influencia a diversidade funcional  $\alpha$ ; e a altura das macrófitas e a distância da margem mais próxima influenciam a diversidade filogenética  $\alpha$ . Nosso estudo também sugere que as distâncias geográficas não influenciam a assembleia de anuros. Concluimos que as assembleias de anuros desse

lago são estruturadas por processos baseados em nicho. Os anuros em geral têm capacidade de dispersão limitada e são afetados pelas características ambientais locais. Quando em habitats fragmentados, essas características podem reduzir sua dispersão, e como as macrófitas podem ser vistas como manchas de habitat para anuros, processos espaciais têm influência limitada neste tipo de ambiente.

Palavras-chave: Amphibia, ecologia de comunidades, lagos amazônicos, processos de nicho, processos neutros

Environmental effects on taxonomic, functional and phylogenetic diversity of anurans in lacustrine macrophyte banks in Central Amazonia

Mahima Hemnani<sup>1</sup>, Pedro Henrique Salomão Ganança<sup>2</sup>, Rubia Neris Machado<sup>3</sup>, Rafael de Fraga<sup>4</sup>, Igor Luis Kaefer<sup>1,2</sup>

<sup>1</sup>Programa de Pós-Graduação em Zoologia, Instituto de Ciências Biológicas, Universidade Federal do Amazonas, Av. General Rodrigo Otávio Jordão Ramos 6200, CEP 69077-000, Manaus, AM, Brazil.

<sup>2</sup>Instituto Nacional de Pesquisas da Amazônia, Programa de Pós-Graduação em Ecologia, Av. André Araújo, 2936, Petrópolis, 69067-375 Manaus, AM, Brazil

<sup>3</sup>Programa de Pós-Graduação em Ciência Animal e Recursos Pesqueiros, Instituto de Ciências Agrárias, Universidade Federal do Amazonas, Av. General Rodrigo Otávio Jordão Ramos 6200, CEP 69077-000, Manaus, AM, Brazil.

<sup>4</sup>Laboratório de Ecologia e Comportamento Animal, Universidade Federal do Oeste do Pará, Rua Vera Paz, s/n, Sale, CEP 68040-255, Santarém, PA, Brazil.

Manuscrito preparado a partir das instruções para autores da revista *Austral Ecology*, exceto pelo posicionamento das figuras que estão ao longo do texto visando facilitar a legibilidade do estudo.

## SUMÁRIO

1. Abstract.....	11
2. Introduction.....	13
3. Methods.....	18
3.1. Study area.....	18
3.2. Sampling design and data collection.....	19
3.3. Anuran sampling.....	19
3.4. Diversity estimates.....	20
3.5. Sampling of environmental variables.....	24
3.6. Data analysis.....	27
4. Results.....	28
5. Discussion.....	32
6. Acknowledgements.....	37
7. Author contributions.....	37
8. Conflict of interest.....	37
9. References.....	38

## **ABSTRACT**

Assemblages can be structured by neutral, niche-based processes, or combinations of both. Understanding the influence of these processes and evaluating diversity measures like taxonomic, functional, and phylogenetic metrics on the structure of ecological assemblages can show patterns and unravel community assembly mechanisms. Anurans (frogs and toads) represent a large part of the diversity of vertebrates in tropical environments and are a key group in assessing diversity patterns at different spatial scales. They can be terrestrial, aquatic and also found in macrophyte banks. These banks or floating meadows are aquatic and semiaquatic rooted or freely floating plants, which provide shelter, feeding, reproduction, spawning and nursery habitats for many organisms, including anurans, and can promote faunal long-distance dispersal and gene flow. Given that macrophyte meadows can influence the structure of anuran's diversity, our study aimed to test the roles of spatial and environmental factors on the taxonomic, functional, and phylogenetic diversity of anuran assemblages found in floating meadows. Our study was held at Catalão lake, Central Amazonia. Sampling on fifty 50 x 5 m plots occurred during the flood period, in macrophyte banks distributed throughout the lake. Geographical and environmental variables were measured at six equidistant points in each plot. We showed that macrophyte height influences taxonomic  $\alpha$  diversity; macrophyte height and nearest margin distance influence taxonomic  $\beta$  diversity; water pH influences functional  $\alpha$  diversity; and macrophyte height and nearest margin distance influence phylogenetic  $\alpha$  diversity. Our study also suggests that geographic distances do not influence the anuran assemblage. We conclude that the anuran assemblages from this lake are niche structured. Anurans in general have limited dispersion ability and are affected by local environmental characteristics. When in fragmented habitats, these characteristics

can reduce their dispersion, and since macrophytes can be seen as habitat patches for anurans, spatial processes have limited influence in this environment.

Key words: Amazonian lakes, Amphibia, community ecology, neutral processes, niche processes

## INTRODUCTION

Elucidating the processes that determine the composition and structure of assemblages is an important challenge in ecology (Jackson *et al.* 2001). The variation in species or trait composition among assemblages can be generated by neutral and niche-based processes (Brown *et al.* 2017), which are related to stochastic and deterministic predictors. Stochastic processes are usually predicted by spatial-temporal variables related to dispersion and local extinctions, while deterministic processes are usually related to competition or species and trait filtering, by which environmental conditions influence the structuring of assemblages (*sensu* Stroud *et al.* 2015). Understanding the influence of these processes at broad (e.g. millions of square kilometers) and regional scales (e.g. thousands to tens of thousands of square kilometers) means unravelling the structuring mechanisms and processes determining assemblages and modifying them over time.

The predictors behind assembly processes can be related to species being able or not to colonize adjacent habitats (Heino *et al.* 2015). Therefore, changes in species composition between assemblages will be mainly related to the geographical distance between samples (Dumbrell *et al.* 2010). This process is related to the Neutral Theory (Hubbel 2001), a model that predicts that the assemblages are grouped randomly, and the patterns of species' co-occurrence are solely due to stochastic processes associated with their dispersion capacity.

Niche processes are related to the Niche Theory, initially proposed by Hutchinson in 1957, which expresses the relationship of species with aspects of the environment, understanding how an organism or assemblage responds to the availability and distribution of resources. The role of niche-based processes in determining assemblages is usually inferred by the influence of the interaction between biotic and abiotic factors,

demonstrating that the change in species or trait composition is related to environmental variables (Jongman *et al.* 1995). Although measuring all niche dimensions is virtually impossible in empirical studies, identifying niche-based processes can generate spatial patterns if species distribution responds to environmental variables. However, when environmental variables are spatially autocorrelated (Bauman *et al.* 2019), distinguishing whether assemblages are structured by environmental variables, geographic distance, or a combination of both is usually puzzling (Dambros *et al.* 2020).

In an assemblage, species diversity may be a measure of both the number of distinct co-occurring species (species richness) and their relative abundances (Starr & Taggart 2006). In this context, environmental variables limiting the distribution of individual species will have the power to define the spatial variation in the assemblage structure (Kraft *et al.* 2015). One of the main processes by which the environment may limit species distribution is trait selection, whereby species with traits better fitted to local or regional environmental conditions are more likely to survive and thrive (Whitfield *et al.* 2012). The understanding of environmental filters selecting species traits has gained some attention in the last decades due to the increasing availability of mathematical models and computer packages with automated functions for their implementation (e.g. Laliberté *et al.* 2010; Pavoine 2020). In addition, if traits carry phylogenetic signals, e.g. when phylogenetically closely related species are functionally equivalent, estimates of phylogenetic diversity can incorporate evolutionary histories in interpreting relationships between environment and assemblage structure (Faith *et al.* 2018).

The environment is, therefore, a selective force acting on species that do not tolerate conditions in a certain location, and changes in the environment (natural or induced) can change species diversity (Naeem 1998). Diversity can be accessed through its multiple facets using functional and phylogenetic measures. These measures differ from those

based on species abundance or occurrence because they incorporate information on species traits or phylogenetic relationships among species, enabling a complementary view to the traditional taxonomic measures of diversity (Cianciaruso *et al.* 2009). Additionally, incorporating functional traits and evolutionary histories into diversity estimates can generate more powerful models to detect assemblage responses to environmental changes (Ricotta *et al.* 2005) and, consequently, increase the explanatory power of environmental factors in the structuring of assemblages (de Campos *et al.* 2018), allowing us to understand a larger portion of the multiplicity of processes involved in the generation and maintenance of diversity patterns (Pavoine *et al.* 2009; Stevens & Tello 2014).

Anurans are a key group in assessing diversity patterns (Jenkins *et al.* 2013) because they usually respond to environmental gradients at different spatial scales even if the environmental variation is subtle (Dias-Terceiro *et al.* 2015; Ferrão *et al.* 2018). They are also important as bioindicators, because they relate intensely with other groups from the local fauna, especially via trophic interactions, and explore multiple habitats within local landscape patches, as most species have both aquatic and terrestrial life stages (Vitt & Caldwell 2013). However, most studies investigating anuran diversity have been conducted in the light of taxonomic diversity (e.g. Rojas-Ahumada *et al.* 2012; Prado & Rossa-Feres 2014), while few studies have assessed functional (e.g. Lescano *et al.* 2018; Leão-Pires *et al.* 2018) or phylogenetic approaches (e.g. Leão-Pires *et al.* 2018). These studies with complementary approaches can, more accurately, show the characteristics of the species or specific clades that make up an assemblage, being useful in determining conservation priorities. The taxonomic, functional, and phylogenetic diversity of anuran assemblages must vary over environmental gradients, and evaluating such variation offers insights into the processes that determine species assembly (Lescano *et al.* 2018).

In the Amazon, anurans can be found in several types of habitats, including upland forests (terra-firme) and wetlands. In wetlands, macrophyte meadows are an important microhabitat for anuran species (Villamarín *et al.* in press). They are herbaceous or grass water plants found on the surface of water bodies in the entire hydrographic basin of the Amazon, usually associated with the white waters of the floodplain forests (Junk 1984). As soon as the water starts to rise, some species grow rapidly vertically, others float and grow horizontally, expanding the surface area in which they occur (Junk 1970; 1997). As water levels begin to recede, currents and rain can dislodge portions of macrophytes to create floating banks that are then transported across the river current (Upton *et al.* 2014). Floating meadows carry associated fauna when lakes are connected to rivers by seasonal rainfall, and consequently cause long-distance dispersal and gene flow, which may limit the condition of rivers as biogeographic barriers (Schiesari *et al.* 2003; Villamarín *et al.* in press), and therefore, can be important for the spatial structure of communities (Fonte *et al.* 2021; Ganança *et al.* 2021). Few studies have investigated the structure of anuran assemblages in aquatic macrophyte meadows in Amazonia, but they have supported the general premise that deterministic factors (i.e. niche-based processes) are more likely to affect assemblage structure (Fonte *et al.* 2021; Ganança *et al.* 2021) than the expected effects of stochastic factors (i.e. neutral processes generating random assemblages) and biogeographic barriers acting at broader scales.

Macrophytes' importance and diversity have been reported for several taxa, such as invertebrates, fish, and amphibians (Schiesari *et al.* 2003; Dias *et al.* 2011; Upton *et al.* 2014). However, studies focusing on the use of macrophytes by anurans are scarce (Upton *et al.* 2014; Ganança *et al.* 2021), and most of them have been carried out in upland forests (e.g., Ribeiro Jr. *et al.* 2012; Rojas-Ahumada *et al.* 2012; Lipinski *et al.* 2020). Anurans, in general, specialize in habitat requirements features (Kanowski *et al.* 2006; Botts *et al.*

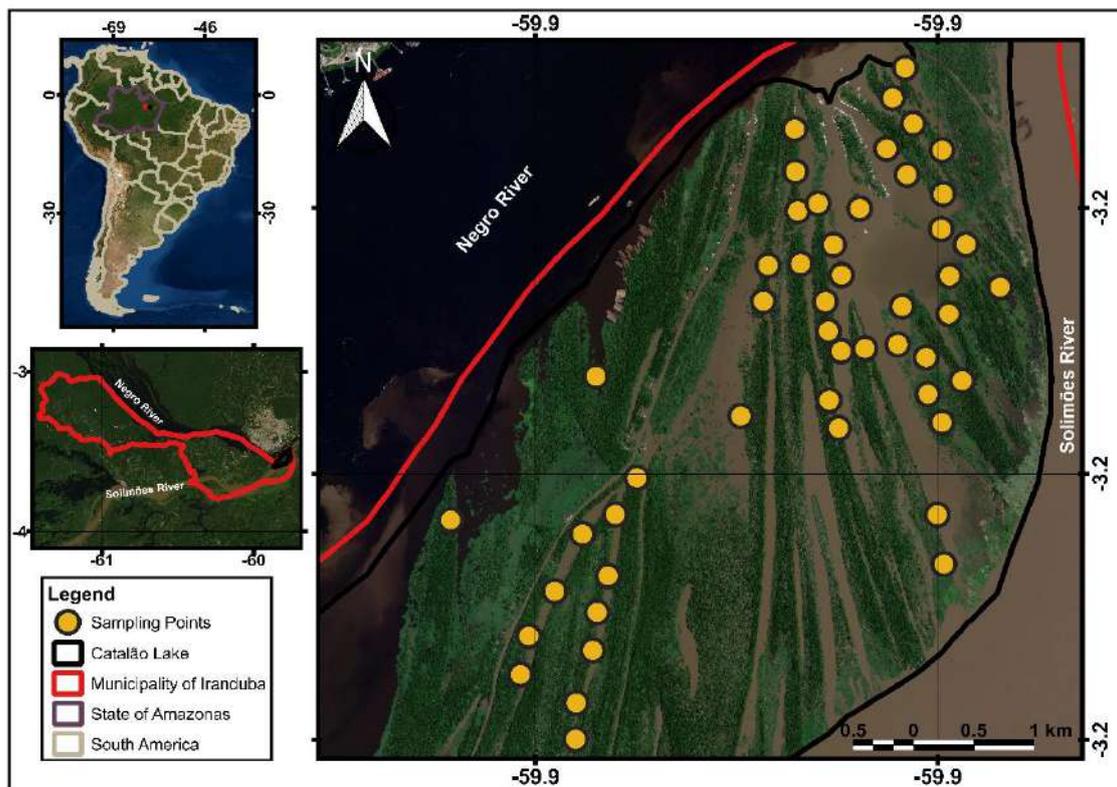
2013), and explore macrophytes for foraging, vocalization, reproduction, as well as egg and tadpole development (Hödl 1977; Schiesari *et al.* 2003; Upton 2015; Villamarín *et al.* in press).

The present study aimed to test the influence of spatial and environmental factors on the taxonomic, functional, and phylogenetic diversity of anuran assemblages found in macrophyte meadows. We tested the hypothesis that environmental variables affect frog diversity estimates because different measures respond to different environmental gradients since they potentially carry complementary information. Specifically, we tested the effects of variables quantifying the structure of macrophyte meadows (plant height and morphotype composition), and surrounding landscapes (water temperature, depth, and pH, dissolved oxygen levels and electrical conductance of the water, air temperature and distance from lake shore) on  $\alpha$  and  $\beta$  diversity estimates based on taxonomic, functional, and phylogenetic data. Investigating multiple assemblage dimensions in a same study system potentially allows accessing broad pictures of factors causing and maintaining biodiversity (Fraga *et al.* 2018).

## METHODS

### Study area

Our study was held at Catalão Lake, municipality of Iranduba, Amazonas, Brazil (Fig. 1). It is a floodplain lake at the confluence of the Amazonas and Negro rivers (Zuchi *et al.* 2020), being periodically flooded by these two rivers (Brito *et al.* 2014). Rio Negro is normally characterized by being poor in electrolytes, with extremely acidic waters, while Rio Solimões is characterized by being rich in electrolytes and biologically as well (Fitkau *et al.* 1975). Although Catalão Lake receives waters from both rivers, its physical and chemical characteristics are influenced by the waters of the Solimões River, with typical characteristics of floodplains, as white water due to the high concentration of sediments from Andean erosion (Brito *et al.* 2014).



**Fig. 1.** Study area in the Catalão lake, located in Central Amazonia, Brazil. Yellow circles are 50 x 5 m sampling units

## **Sampling design and data collection**

The sampling was carried out from 15th February until 13th March 2020, during the flood period, in macrophyte meadows distributed throughout the lake. Fifty plots, 50 m long and 5 m wide were sampled once. Although temporal repetitions could increase detection probabilities of some species, repeating surveys in the same sites where plots were installed is challenging, because portions of macrophyte meadows are often detached, and move away. However, since our sampling was conducted during the reproductive peak of most species, we reduced any detectability effect in our models by being able to detect most species through male vocalization. Neighbouring plots had a minimum distance of 300 m (adapted from Ganança *et al.* 2021). We used a 10-m long boat to sample frog occurrences and environmental variables for each plot for about 1 h between 18h30 and 22h00. All plots were georeferenced so that we could calculate Euclidean distances between them.

## **Sampling of environmental variables**

We measured environmental variables that are associated with vegetation cover, plant stratification and physicochemical parameters of the water. These variables were measured every 10 m within plots (6 values per plot) and we used mean values per plot for analysis. We summarised all environmental gradients measured in Table 2.

We measured the physicochemical properties of the water because they can guide the selection of oviposition sites by some species, in addition to affecting the development of embryos and larvae (Touchon & Worley 2015). We measured dissolved oxygen (mg/L), pH, electrical conductivity ( $\mu\text{s}\cdot\text{cm}^{-1}$ ), air and water temperature ( $^{\circ}\text{C}$ ) and depth of the water column, respectively using an oximeter AKSO model AK84, a pH meter AKSO/AK86, conductivity meter AKSO/AK83 (for electrical conductivity and water and

air temperature), and a graduated tape with a weight attached to its end. The measurements, except the water column, were taken on the water surface (5 cm deep). Only one person was responsible for collecting all the environmental variables during the entire sampling. We collected these variables at night to reduce the effects of variation on water physicochemical parameters throughout the day.

We measured vegetation cover across macrophyte meadows because this variable potentially filters species, traits, or clades by limiting the availability of foraging, calling, and breeding sites. Additionally, vegetation cover can cause spatial variation in the composition of anuran assemblages along macrophyte meadows, because while some plants form good dispersal corridors, others are better suited as shelters to hide from predators (Ganança *et al.* 2021). To quantify vegetation cover we randomly established a 2 m<sup>2</sup> area within each plot, where we measured average plant height and the proportional occurrence of morphotypes. We classified morphotypes based on the shape and size of the stem, leaves and roots, according to a field guide for aquatic macrophytes (Demarchi *et al.* 2018), and summarized the values of morphotype occurrence using PCA (Principal Component Analysis). We used the PCA axis 1, which captured 51% of the variance of morphotype frequency.

We also measured the distance from each plot to the nearest margin with a laser gauge (RangeFinder Handheld PD-58N). This variable can be an important predictor of the structure of anuran assemblages because the ability to colonize macrophyte banks far from the margin depends on the ability to disperse over plants or into water, which in turn is deeply associated with some species traits, such as body size, and the presence of toe pads.

**Table 2.** Minimum (Min), maximum (Max), mean and standard deviation (SD) values of environmental variables measured in 50 plots distributed along the Catalão Lake, Central Amazonia. Vegetation cover is not in the table because it is a PCA derived variable

Gradient	Min	Max	Mean	Sd
Macrophyte height (cm)	1.54	61.58	30.39	16.144
pH	3.88	6.92	5.772	0.691
Dissolved oxygen (mg/ml <sup>3</sup> )	4	70.37	31.122	15.482
Air temperature	24.67	30.2	27.583	0.997
Water temperature (°C)	20.97	33.23	30.264	2.007
Electrical conductivity (µs.cm <sup>-1</sup> )	10.16	122.1	59.616	30.03
Water depth (m)	0.89	5.43	2.948	1.061
Distance from the bank to the nearest border (m)	5.55	187.41	38.859	37.414

### **Anuran sampling**

We sampled anurans simultaneously using visual encounters and acoustic surveys (Crump & Scott 1994; Zimmerman 1994). These methods are complementary because species with low visual detectability such as those with small bodies or cryptic colours can be detected by calling males. The nomenclature of taxonomic categories follows Frost (2020). We collected a maximum of five voucher specimens per species, which were killed using 5% lidocaine, fixed in 10% formaldehyde, and kept in 70% alcohol in the Zoological Collection at the Federal University of Amazonas. All collection procedures were authorized by the environmental authority through the permit SISBIO number 72521-1.

### **Diversity estimates**

#### Taxonomic diversity

To quantify anuran  $\alpha$ -diversity, we used the Fisher's index, which assumes that species abundance follows a log-series distribution (Fisher *et al.* 1943). We used this index because, compared to other indices such as Simpson, it does not overemphasize common or rare species, which could cause diversity estimates to be unstable over time or space (Kempton & Taylor 1976). We estimated  $\beta$ -diversity using pairwise dissimilarities in assemblage composition among plots, which we estimated using Non-Metric Multidimensional Scale (NMDS) with the Bray-Curtis index applied to abundance data. Since NMDS axes are not orthogonal, and therefore not independent, we configured it to reduce the dimensionality of species abundance in one dimension ( $k = 1$ ). The best solution returned by the NMDS was 87% related to the original distances (Stress = 0.35),

which indicates that it is a reliable univariate representation of the multivariate species abundance since little information has been lost by reducing dimensionalities.

### Functional diversity

To estimate functional diversity, we relied on seven traits which are potentially filtered by local environmental conditions varying across plots. We measured morphometric traits because they indicate dispersal and competition abilities, as well as feeding habits. These traits are thought to also affect the occupancy of macrophytes by determining the ability to climb and disperse over the vegetation cover and water (Ganança *et al.* 2021).

We measured one to five individuals of each species, using a digital caliper. For the species for which we found less than five individuals, we supplemented our data with published information (Schalk *et al.* 2014; Upton *et al.* 2014; Lescano *et al.* 2018). The functionalities of each trait can be found in Table 1.

**Table 1.** List of functional traits and their ecological meanings. CO = continuous, CA = categorical, OB = observed, LT = literature

Functional trait	Trait definition	Functional meaning	Type of data	Source
Snout-vent length (SVL)	Maximum length (snout-vent length) of post metamorphic stages	Acquisition of resources and type of vegetation in which they can be found. Measured in mm	CO	OB
Head length (HD) and Head width (HW)	HD is the length of the head measured in full-face view and HW is the maximum	Anuran diet (resource acquisition), since the size and type of	CO	OB

	width of the head in the full-face view	food items consumed by amphibians can be influenced by the shape of the skull and body size (Schalk <i>et al.</i> 2014). Measured in mm.		
Tibia-fibula relative length and total length of the posterior limb	Tibia-fibula/ hind-limb length ratio	Habitat use (Lescano <i>et al.</i> 2018). High values indicate a lifestyle in plants with greater heights, due to the ease of jumping and occupying these plants (Upton <i>et al.</i> 2014). Measured in mm.	CO	OB
Body mass	The individual mass measured in grams (g)	Type of vegetation in which it can be found.	CO	OB
Oviposition site	Egg's deposition location (water or not)	To know if the eggs deposition is on the macrophytes or in the water	CA	LT
Toe pads	Toe pads on the ends of the fingers and toes and leap from branch to branch or	Climbing structures that reflect arboreal habitats	CA	OB

	from leaf to leaf (present or not)			
Tadpole microhabitat	Position of the tadpoles the water column (benthic or not)	Breeding biology and larval development	CA	LT

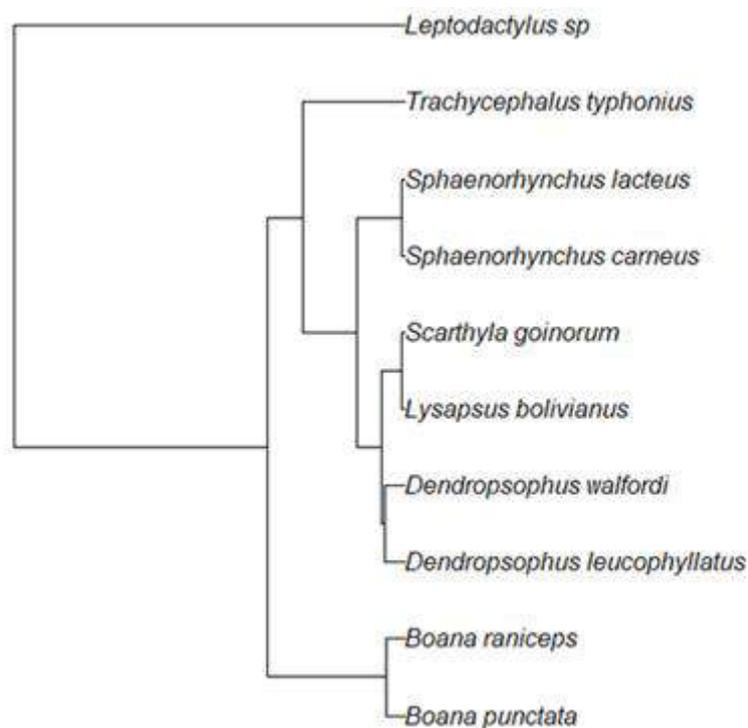
We calculated Gower distances between species using the `gowdis` function of the FD R-package (Laliberté & Legendre 2010; Laliberté *et al.* 2014). The Gower distance calculated by the FD package transforms all traits to vary on the same scale, so that continuous and categorical traits can be analysed from a single matrix (Fraga *et al.* 2018). In this study, we represented functional diversity by the functional richness index (FRic), because it represents the amount of functional space filled by a given assemblage, which is estimated by the convex hull volume of a polygon formed by the distribution of traits weighted by species abundance (Villéger *et al.* 2008).

We also carried out a fourthcorner model to identify the association between environmental factors and functional traits (Legendre *et al.* 1997). This approach was useful to estimate how each trait is affected by each environmental variable individually. We implemented a fourthcorner model using the `mvaabund` R-package (Wang *et al.* 2012).

#### Phylogenetic diversity

To represent the evolutionary relationships between species from the Catalão Lake we followed the phylogenetic hypothesis proposed by Jetz & Pyron (2018). This hypothesis was based on ten nuclear and five mitochondrial genes and was statistically well supported. To obtain a phylogenetic sub-tree with the species we found in the Catalão Lake, we extracted the consensus tree from Jetz & Pyron (2018) in the Phylocom program (Webb *et al.* 2008) and removed the external group (*Homo sapiens*) and unsampled

species (Fig. 2.) using the `drop.tip` function from the `ape` R-package (Paradis & Schliep 2019). Then, we used the `phylosor` function of the `stats` R-package (R core team 2019) to extract the phylogenetic distance matrix from the sub-tree and summarized it as a Principal Coordinates axis (PCoA). Through this approach, the estimated values express branch lengths of the phylogenetic tree that are shared between paired plots. Therefore, the higher the values obtained, the closer the phylogenetic relationship between the species that compose assemblages.



**Fig. 2.** Phylogenetic tree showing the genetic relationship between anurans found at the Catalão Lake (adapted from Jetz & Pyron 2018)

## Data analysis

### Effect of environmental variables on diversity metrics

All analyses were performed in the R computational environment (R Core Team 2020). To test the effect of environmental variables on anuran's diversity metrics, we

used multiple linear models, which were setup with diversity estimates as response variables and environmental gradients as predictors. We applied a stepwise regression on the multiple linear models to select the most parsimonious variable subset so that the less informative predictor variables were sequentially removed from the full model, and a final model was selected by the lowest Akaike Information Criterion (AIC) value. Our multiple linear models tested in this study were validated by Shapiro-Wilk tests to evaluate if the residuals did not differ from a normal theoretical distribution. We estimated the variance inflation factor (VIF) using the car R-package (Fox & Weisberg 2011) to make sure that none of the final models kept highly redundant predictor variables.

#### Effect of geographical distance on diversity metrics

We used Moran's I global tests and partial Mantel tests, with the geographic distance matrix and the environmental variable that best explained each diversity measure. These tests were useful in making sure that any effect of environmental variables on frog diversity estimates were not caused by geographical distances among plots instead of the environmental variables themselves (Haining 2001).

## **RESULTS**

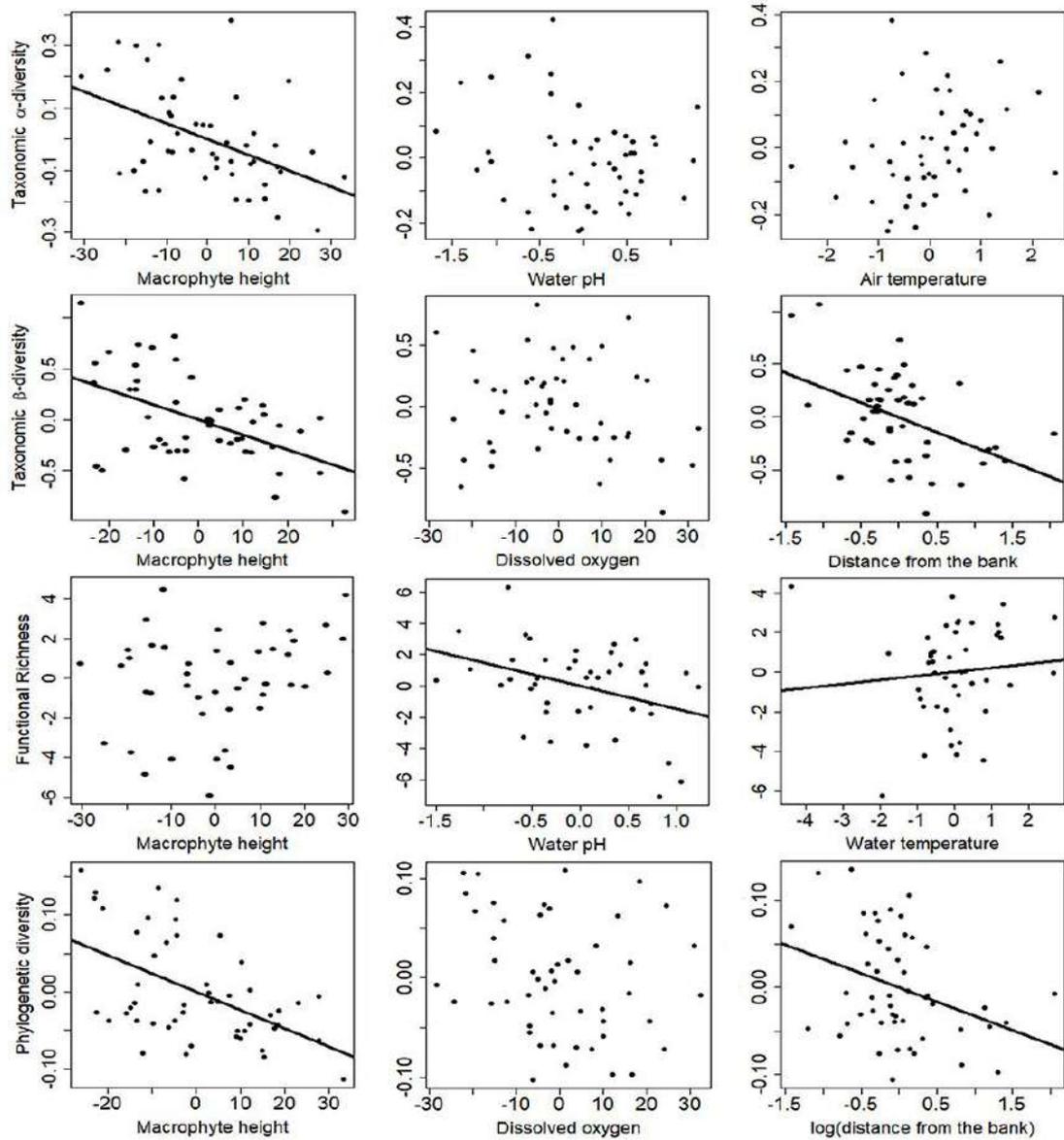
We recorded 1748 individual anurans distributed in 10 species and two families (Table 3). The most frequent species sampled was *Dendropsophus walfordi* (Bokermann, 1962), and the less frequent species was *Trachycephalus typhonius* (Linnaeus, 1758).

**Table 3.** Frog species found in the 50 plots distributed at the Catalão Lake, Central Amazonia

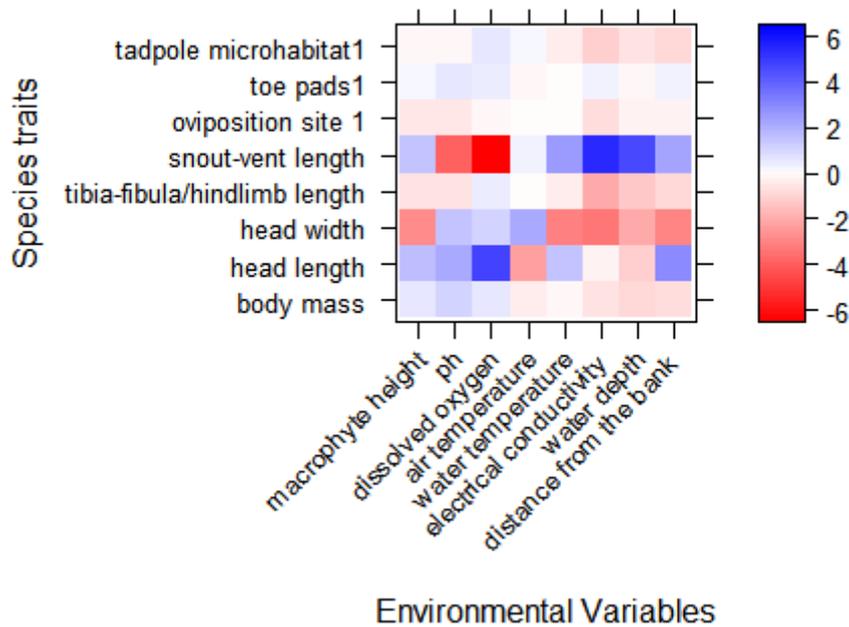
Family/Species	Abundance (number of individuals)	Occurrence frequency (%)
<b>Hylidae</b>		
<i>Boana punctata</i> (Schneider, 1799)	5	0.29
<i>Boana raniceps</i> (Cope, 1862)	117	6.67
<i>Dendropsophus gr leocophyllatus</i> (Beireis, 1783)	28	1.60
<i>Dendropsophus walfordi</i> (Bokermann, 1962)	658	37.51
<i>Lysapsus aff. bolivianus</i> (Gallardo, 1961)	627	35.75
<i>Scarthyla goinorum</i> (Bokermann, 1962)	1	0.06
<i>Sphaenorhynchus lacteus</i> (Daudin, 1800)	18	1.03
<i>Sphaenorhynchus carneus</i> (Cope, 1868)	10	0.57
<i>Trachycephalus typhonius</i> (Linnaeus, 1758)	1	0.06
<b>Leptodactylidae</b>		
<i>Leptodactylus</i> sp	289	16.48

Regarding taxonomic  $\alpha$ -diversity, only macrophyte height showed a negative effect ( $p < 0.01$ ,  $t = -3.48$ ,  $SE < 0.01$ ). For taxonomic  $\beta$ -diversity, the environmental influence was determined by macrophyte height ( $p < 0.01$ ,  $t = 3.878$ ,  $SE < 0.01$ ), and nearest margin distance ( $p < 0.01$ ,  $t = 3.25$ ,  $SE = 0.08$ ). For functional diversity, the only predictor was water pH ( $p = 0.01$ ,  $t = -2.51$ ,  $SE = 0.58$ ). For phylogenetic diversity, the environmental influence was determined by macrophyte height ( $p < 0.01$ ,  $t = -4.1$ ,  $SE < 0.01$ ), and nearest margin distance ( $p = 0.01$ ,  $t = -2.51$ ,  $SE = 0.01$ ). VIF values ranged between 1.18 and 2, so no model was biased by multicollinearity among the predictor variables. Graphic representations of partials derived from all models selected stepwise can be found in Fig. 3.

The stronger correlations returned by the fourthcorner analysis showed that body size is positively related to electric conductivity in the water and water depth, and negatively related to water dissolved oxygen and pH. Additionally, body size and head length have been positively related to distance from the bank, showing that larger anurans have been able to colonize sites far from the margins. Head length was negatively associated with macrophyte height. These findings show a combination of physical-chemical properties of water limiting the presence of some species being able to disperse away from the lake shore and colonizing some sites that can affect the development of the tadpoles. A colour coding of the fourthcorner analysis results can be found in Fig. 4



**Fig. 3.** Partials derived from multiple linear models tested to quantify the effects of environmental variables on frog  $\alpha$ -diversity (A, B, C),  $\beta$ -diversity (D, E, F), functional diversity (G, H, I) and phylogenetic diversity (J, K, L) estimates. The more informative environmental variables for each metric (based on the stepwise regression) are in the image.



**Fig. 4.** Fourth corner interaction between amphibian species traits and environmental variables. Positive and negative associations are shown respectively in blue and red.

Moran's I global test showed that the residuals are not spatially autocorrelated ( $p = 0.07$ ), and the partial Mantel test was not able to reject the null hypothesis ( $p > 0.05$  in all cases). The results suggest that our models are not biased by spatial autocorrelation, and the results can be interpreted considering environmental filtering structuring assemblages. Overall, our models indicated that anuran assemblages from the Catalão Lake macrophyte meadows are more influenced by deterministic than neutral factors.

## DISCUSSION

Our results showed that anuran assemblages from aquatic macrophyte meadows in Central Amazonia are more structured by environmental filtering limiting species local distribution than geographic distance limiting dispersal, differently from a large-scale study showing that amphibian diversity in floating meadows is mostly determined by stochastic (i.e. random/neutral) processes (Fonte *et al.* 2021). Overall, our results support niche-based processes as more powerful predictors of biodiversity than purely randomly based neutral processes. Environmental filtering has been widely demonstrated as a major source of spatial variation in local anuran assemblages, but usually in forests (Ribeiro-Jr. *et al.* 2012; Rojas-Ahumada *et al.* 2012; Dias-Terceiro *et al.* 2015). Here we showed that distinct assemblages on distinct portions of environmental gradients can be expected even in habitats that are locally rare (Ganança *et al.* 2021), and that are rarely considered to be distinct from surrounding forests, even though they cover enough environmental heterogeneity to contain multiple assemblages (Fonte *et al.* 2021; Villamarín *et al.* in press).

Vegetation cover is often reported as an important predictor of anuran richness and composition, because the more heterogeneous the vegetation, the greater the variety of habitats that can be occupied by different species (Pearman 1997; Ernst & Rodel 2005, Ernst *et al.* 2006). Additionally, in macrophyte meadows sites with high vegetation provide higher levels of vertical stratification, which can increase the number of co-occurring species (Ganança *et al.* 2021). However, the relationship between macrophyte height and Fisher's  $\alpha$ -diversity was unexpectedly negative in the Catalão Lake, contrary to what Ganança *et al.* (2021) found in a lake near the lower Amazon River. The inconsistency between both studies can be explained by the differences in the vegetation structure between the lakes, as the average height of the vegetation cover was 1.8 times

lower in Catalão than in the lower Amazon River. Alternatively, most of the species from the regional pool that we found in Catalão have a biological affinity with habitats close to water. In any case, our results encourage the empirical replication of these studies in other vegetated water bodies, since it is currently impossible to predict ecological relationships that should be generalized.

Although our results on  $\alpha$ -diversity were opposed to a lake in the lower Amazon River (Ganança *et al.* 2021), they converged on identifying macrophyte height as a source of  $\beta$ -diversity. Although plots with tall vegetation had fewer species, some species seem specialized in occupying these environments, which ultimately has resulted in assemblage turnover along the gradient of vegetation height. Such specialization may be an evolutionarily fixed trait, as macrophyte height also significantly affected our estimate of phylogenetic diversity. However, the effects of macrophyte height on the spatial variation in assemblage composition is hardly mediated by ecomorphological relationships, such as through species with toe pads climbing tall plants. This claim is supported by the fact that we found no evidence of macrophyte height affecting estimates of functional diversity or occurrence of individual traits.

Functional diversity was affected by pH, which is possibly related to non-random selection of oviposition sites. Physical and chemical water parameters can affect the growth and survival of many anuran species (Duellman & Trueb 1994) and limit the occupation of habitats by some species (Jorge *et al.* 2016). Therefore, the physicochemical quality of water potentially affects the breeding, hatching, development, locomotion, mortality, and habitat distributions of species (Thabah *et al.* 2018; Shangpliang *et al.* 2018), especially in the early developmental stages, when amphibians can be severely affected by low pH as it affects their ionic regulation (Vignoli *et al.* 2007).

Although water depth, distance from the bank and electrical conductivity of the water did not affect the estimated functional richness, the fourthcorner analysis showed evidence that these variables may affect individual traits. For instance, we found greater chances of finding species with large bodies and heads in plots far from the bank, inserted in deep water. These findings are potentially related to the fact that larger anurans have a greater ability to disperse (Searcy *et al.* 2018). Additionally, we found that plots installed in sites with higher electrical conductivity of water harbor larger species. This finding may be related to the fact that Catalão Lake receives a greater charge of electrolytes from Andean erosion carried by the Amazon River during the peak of floods (Brito *et al.* 2014). Greater availability of nutrients possibly feeds richer trophic networks, which increases the chances of survival of relatively large animals. Alternatively, high electrical conductivity may reflect pollution from sewage, and larger frogs are more tolerant of environmental disturbances (Brady *et al.* 2018). Ultimately, the fourthcorner results show that estimating multivariate functional diversity can overlook important environmental effects on trait selection, which can only be detected on individual traits.

The estimated phylogenetic diversity was influenced by macrophyte height and distance from the bank. This means that, farther from the margin, the species are phylogenetically more similar to each other. According to Ribeiro *et al.* 2017, anuran biomass decreases with increasing distance to the closest waterbody. As mentioned before, anurans have low dispersal ability, hence, colonizing some isolated meadows can be impossible for some species (Silva *et al.* 2011). Other studies have also found that anuran diversity has been determined as a response to either distinct types of vegetation, distance to water bodies or environmental heterogeneity (Goncalves, Crivellari, & Conte 2015; Ribeiro *et al.* 2017).

Our findings also show that stochastic processes have little influence on anurans assembly. It has been predicted that organisms with limited dispersal ability, due to both intrinsic (e.g. natural low movement capacity, willingness to move and body size) and/or extrinsic traits (e.g. imposed by landscape effects), would be prevented from colonising sites that are further apart (Delatorre *et al.* 2015). Anurans, in general, depend on environment and have low dispersal ability (Smith & Green 2005). Most of the studies on anurans have been conducted in forests (Prado & Rossa Feres 2014; Leão-Pires *et al.* 2018). Anuran beta diversity in this type of study region may also be driven by stochastic mechanisms, which supposes that population dynamics is not dependent on environmental parameters but regulated by ecological drift and/or random dispersal (Hubbel 2001; Leão-Pires *et al.* 2018).

In conclusion, the present study showed the role of environmental conditions shaping anuran assembly and that different biodiversity metrics are influenced by different environmental gradients, as they carry complementary information on biodiversity. We specifically showed that in macrophyte meadows in central Amazonia, niche-based processes are better predictors of assemblage richness and composition than dispersal-based neutral processes that are important at larger scales (Fonte *et al.* 2021). A multi-faceted assessment of diversity usually improves our understanding of the mechanisms underlying biodiversity patterns and community assembly (Borges *et al.* 2021) and is essential to direct conservation efforts beyond pure species numbers, towards a more inclusive approach prioritizing ecosystem function and stability (Cadotte *et al.* 2011). For instance, in a single study system we show that although apparently homogeneous, the aquatic vegetation of a tropical lake contains high enough environmental heterogeneity to generate biotic complementarities between sites. Biotic

complementarities usually provide a solid basis for conservation because it allows for an assessment of the degree of uniqueness of biodiversity.

## **ACKNOWLEDGEMENTS**

We thank Marcelo Menin (*in memoriam*) for his immense contribution towards the idea, execution, and valuable suggestions for this project. We are grateful to Mauro Coelho Filho, our guide and support during field activities; Instituto Nacional de Pesquisas da Amazônia (INPA) which granted access to the Catalão Lake accommodations; Brazilian Sistema de Autorização e Informação em Biodiversidade - SISBIO for collection permits (# 72521-1). This study was financed by the Brazilian Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) through the project "Anfíbios da Amazônia Brasileira: taxonomia, distribuição e ecologia" (Process # 405640/2016-1), and by Brazilian Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES). Fundação de Amparo à Pesquisa do Estado do Amazonas granted MH a master's degree scholarship.

## **AUTHOR CONTRIBUTIONS**

All authors contributed to the study conception and design. Data collection in the field was performed by Mahima Hemnani, Pedro Henrique Salomão Ganança, Rubia Neris Machado and Igor Kaefer. The analyses were performed by Mahima Hemnani and Rafael de Fraga. The first draft of the manuscript was written by Mahima Hemnani, and all authors contributed on subsequent versions by reviewing and editing the manuscript. All authors read and approved the final manuscript.

## **CONFLICT OF INTEREST**

The authors declare no conflict of interest.

## REFERENCES

Barth B. J. & Wilson R. S. (2010) Life in acid: interactive effects of pH and natural organic acids on growth, development, and locomotor performance of larval striped marsh frogs (*Limnodynastes peronii*). *J. Exp. Biol.* **213(8)**, 1293-1300.

Bauman D., Vleminckx J., Hardy O. J. & Drouet, T. (2019) Testing and interpreting the shared space-environment fraction in variation partitioning analyses of ecological data. *Oikos*. **128(2)**, 274-285.

Borges S. H., do RS Tavares, T., Crouch, N. M. & Baccaro, F. (2021) Sucessional trajectories of bird assemblages in Amazonian secondary forests: Perspectives from complementary biodiversity dimensions. *For. Ecol. Manag.* **483**, 118731.

Botts E., Erasmus B. & Alexander, G. (2013) Small range size and narrow niche breadth predict range contractions in South African frogs. *Glob. Ecol. Biogeogr.* **22**, 567–576.

Brady, S. P., Zamora-Camacho, F. J., Eriksson, F. A., Goedert, D., Comas, M. & Calsbeek, R. (2019). Fitter frogs from polluted ponds: The complex impacts of human-altered environments. *Evol. Appl.* **12(7)**, 1360-1370.

Brito, J. G. D., Alves, L. F. & Espírito Santo, H. M. V. (2014) Seasonal and spatial variations in limnological conditions of a floodplain lake (Lake Catalão) connected to both the Solimões and Negro Rivers, Central Amazonia. *Acta Amaz.* **44**, 121-133.

Brito, J. G. D., Alves, L. F., Santo, E. & Viana, H. M. (2014) Seasonal and spatial variations in limnological conditions of a floodplain lake (Lake Catalão) connected to both the Solimões and Negro Rivers, Central Amazonia. *Acta Amazon.* **44(1)**, 121-133.

Brown, B.L., Sokol, E.R., Skelton, J. & Tornwall, B. (2017) Making sense of metacommunities: dispelling the mythology of a metacommunity typology. *Oecologia*. **183**, 643-652.

Cadotte, M. W., Carscadden, K., & Mirotchnick, N. (2011) Beyond species: functional diversity and the maintenance of ecological processes and services. *J Appl Ecol*. **48(5)**, 1079-1087.

Cienciaruso, M. V., Silva, I. A. & Batalha, M. A. (2009) Diversidades filogenética e funcional: novas abordagens para a ecologia de comunidades. *Biota Neotrop*. **9(3)**, 93-103.

Couto, A. P., Ferreira, E., Torres, R. T., & Fonseca, C. (2017) Local and landscape drivers of pond-breeding amphibian diversity at the northern edge of the Mediterranean. *Herpetologica*. **73(1)**, 10-17.

Dambros, C., Zuquim, G., Moulatlet, G. M., Costa, F. R., Tuomisto, H., Ribas, C. C. & Magnusson, W. E. (2020) The role of environmental filtering, geographic distance and dispersal barriers in shaping the turnover of plant and animal species in Amazonia. *Biodivers. Conserv*. **29(13)**, 3609-3634.

de Campos, R., Lansac-Tôha, F. M., da Conceição, E. D. O., Martens, K. & Higuti, J. (2018) Factors affecting the metacommunity structure of periphytic ostracods (Crustacea, Ostracoda): a deconstruction approach based on biological traits. *Aquat. Sci*. **80(16)**, 1-12

de Fraga, R., Ferrão, M., Stow, A. J., Magnusson, W. E. & Lima, A. P. (2018) Different environmental gradients affect different measures of snake  $\beta$ -diversity in the Amazon rainforests. *Peer J*. **6**, e5628.

Delatorre, M., Cunha, N., Raizer, J., & Ferreira, V. L. (2015) Evidence of stochasticity driving anuran metacommunity structure in the Pantanal wetlands. *Freshw. Biol.* **60(11)**, 2197-2207.

Demarchi, L. O.; Cruz, J. D.; Junk, W.; Lopes, A.; Schöngart, J.; Wittmann, F. & Piedade, M. T. F. (2018) Guia de campo de herbáceas aquáticas: várzea Amazônica. Editora do INPA, Manaus.

Dias, M. S., de Toledo, J. J., Jardim, M. M., de Figueiredo, F. O. G., de Oliveira Cordeiro, C. L., Gomes, A. C. S. & Zuanon, J. (2011) Congruence between fish and plant assemblages in drifting macrophyte rafts in Central Amazonia. *Hydrobiologia.* **661(1)**, 457-461.

Dias-Terceiro, R. G., Kaefer, I. L., de Fraga, R., de Araújo, M. C., Simões, P. I., & Lima, A. P. (2015) A matter of scale: historical and environmental factors structure anuran assemblages from the Upper Madeira River, Amazonia. *Biotropica.* **47(2)**, 259-266.

Dumbrell, A. J., Nelson, M., Helgason, T., Dytham, C. & Fitter, A. H. (2010) Relative roles of niche and neutral processes in structuring a soil microbial community. *ISME J.* **4(3)**, 337.

Ernst, R., & Rödel, M. O. (2005) Anthropogenically induced changes of predictability in tropical anuran assemblages. *Ecology.* **86(11)**, 3111-3118.

Ernst, R., Linsenmair, K. E., & Rödel, M. O. (2006) Diversity erosion beyond the species level: dramatic loss of functional diversity after selective logging in two tropical amphibian communities. *Biol. Conserv.* **133(2)**, 143-155.

Faith, D. P. (2018) Phylogenetic diversity and conservation evaluation: perspectives on multiple values, indices, and scales of application. In *Phylogenetic diversity* (pp. 1-26). Springer, Cham.

Ferrão, M., de Fraga, R., Moravec, J., Kaefer, I. L., & Lima, A. P. (2018) A new species of Amazonian snouted treefrog (Hylidae: *Scinax*) with description of a novel species-habitat association for an aquatic breeding frog. *PeerJ*. **6**, e4321.

Fisher R. A., Corbet A. S. & Willians C. B. (1943) The relation between the number of species and the number of individuals in a random sample of an animal population. *J. Anim. Ecol.* **12**, 42–58.

Fitkau, E. J., Irmeler, U., Junk, W. J., Reiss, F. & Schmidt, G. W. (1975) Productivity, biomass and populations dynamics in amazon water bodies. P.289-311. In: E. Golley E E. Medina (Eds.). *Tropical Ecological Systems, Trends in Terrestrial and Aquatic Research*. Springer-Verlag, New York,

Flynn, D. F. B., Gogol-Prokurat, M., Nogeire, T., Molinari, N., Richers, B. T., Lin, B. B. & DeClerck, F. (2009) Loss of functional diversity under land use intensification across multiple taxa. *Ecol Lett.* **12**, 22–33.

Fonte, L. F. M. D., Latombe, G., Gordo, M., Menin, M., de Almeida, A. P., Hui, C. & Lötters, S. (2021) Amphibian diversity in the Amazonian floating meadows: a Hanski core-satellite species system. *Ecography*. <https://doi.org/10.1111/ecog.05610>

Fox J. & Weisberg S. (2011) *An R Companion to Applied Regression*, 2nd edn. SAGE Publications Inc, Thousand Oaks.

Frost D. R. (2020) Amphibian species of the world: an online reference. Version 6.0. American Museum of Natural History, New York. Available from URL: <http://research.amnh.org/herpetology/amphibia/index.html>

Ganança, P. H. S., Santos Jr, A. P., Kawashita-Ribeiro, R. A., de Vasconcelos Neto, L. B., dos Santos Júnior, I. A., Guedes, D. D. S., & de Fraga, R. (2021) Habitats determining local frog assemblages within aquatic macrophyte meadows in Amazonia, through species traits filtering. *Austral Ecol.* **46(4)**, 574-587.

Goncalves, D. S., Crivellari, L. B., & Conte, C. E. (2015) Linking environmental drivers with amphibian species diversity in ponds from subtropical grasslands. *Anais da Academia Brasileira de Ciências.* **87**, 1751-1762.

Göthe, E., Baattrup-Pedersen, A., Wiberg-Larsen, P., Graeber, D., Kristensen, E. A. & Friberg, N. (2016) Environmental and spatial controls of taxonomic versus trait composition of stream biota. *Freshw. Biol.* **62**, 397–413.

Heino, J., Melo, A. S., Siqueira, T., Soininen, J., Valanko, S. & Bini, L. M. (2015) Metacommunity organisation, spatial extent and dispersal in aquatic systems: patterns, processes, and prospects. *Freshw. Biol.* **60(5)**, 845-869.

Hödl, W. R. (1977) Call differences and calling site segregation in anuran species from Central Amazonian floating meadows. *Oecologia*, **28**: 351-363.

Hubbell, S.P. (2001) *The Unified Neutral Theory of Biodiversity and Biogeography*. Monographs in Population Biology. Princeton University Press: Princeton, 346p.

Hutchinson, G. E. (1957) *A Treatise on Limnology*, volume II. John Wiley & Sons: New York, London & Sidney, 1093p.

Jackson, D. A., Peres-Neto, P. R. & Olden, J. D. (2001) What controls who is where in freshwater fish communities – the roles of biotic, abiotic, and spatial factors. *Can. J. Fish. Aquat. Sci.* **58**, 157-170.

Jenkins, C. N., Pimm, S. L. & Joppa, L. N. (2013) Global patterns of terrestrial vertebrate diversity and conservation. *PNAS USA.* **110(28)**, 2602-2610.

Jetz, W. & Pyron, R. A. (2018) The interplay of past diversification and evolutionary isolation with present imperilment across the amphibian tree of life. *Nat. Ecol. Evol.* **2(5)**, 850-858.

Jongman, E. & Jongman, S. R. R. (1995) Data analysis in community and landscape ecology. Cambridge university press, 300p.

Junk, W. (1970) Investigations on the ecology and production biology of the floating meadows (*Paspalo echinochloetum*) on the middle Amazon, part 1, the floating vegetation and its ecology. *Amazoniana.* **2**, 449-495.

Junk, W. (1997) The Central Amazon Floodplain: Ecology of a Pulsing System. Springer, Berlin, Hiedelberg, 556p.

Junk, W. J. & Howard-Williams, C. (1984) Ecology of aquatic macrophytes in Amazonia. In: *The Amazon*. Springer, Dordrecht, p. 269-293.

Kanowski, J. J., Reis, T. M., Catterall, C. P. & Piper, S. D. (2006) Factors affecting the use of reforested sites by reptiles in cleared rainforest landscapes in tropical and subtropical Australia. *Restor. Ecol.* **14(1)**, 67-76.

Kempton R. A. & Taylor L. R. (1976) Models and statistics for species diversity. *Nature.* **262**, 818-20.

Knauth, D. S., Pires, M. M., Stenert, C. & Maltchik, L. (2019) Disentangling the role of niche-based and spatial processes on anuran beta diversity in temporary ponds along a forest–grassland transition. *Aquat. Sci.* **81(63)**, 1-13.

Kraft, N. J., Adler, P. B., Godoy, O., James, E. C., Fuller, S. & Levine, J. M. 2015. Community assembly, coexistence and the environmental filtering metaphor. *Funct. Ecol.* **29(5)**, 592-599.

Laliberte E., Legendre P. & Shipley B. (2014) FD: measuring functional diversity from multiple traits, and other tools for functional ecology. R package version 1.0-12.

Laliberté, E. & Legendre, P. (2010) A distance-based framework for measuring functional diversity from multiple traits. *Ecology*. **91(1)**, 299-305.

Leão-Pires, T. A., Luiz, A. M. & Sawaya, R. J. (2018) The complex roles of space and environment in structuring functional, taxonomic and phylogenetic beta diversity of frogs in the Atlantic Forest. *PloS One*. **13(4)**, e0196066.

Legendre, P. & Legendre, L. F. (2012) *Numerical ecology*. 3rd ed. Elsevier, United Kingdom, 989p.

Lescano, J. N., Miloch, D. & Leynaud, G. C. (2018) Functional traits reveal environmental constraints on amphibian community assembly in a subtropical dry forest. *Austral Ecol.* **43(6)**, 623-634.

Lipinski, V. M., Schuch, A. P. & dos Santos, T. G. (2020) Changes on Anuran Tadpole Functional Diversity along an Environmental Gradient at the Southernmost Atlantic Rainforest Remnant. *Asian Herpetol. Res.* **11(1)**, 63-70.

McGill, B. J., Enquist, B. J., Weiher, E. & Westoby, M. (2006). Rebuilding community ecology from functional traits. *Trends Ecol. Evol.* **21(4)**, 178-185.

Naeem, S. (1998) Species redundancy and ecosystem reliability. *Conserv. Biol.* **12** (1), 39-45.

Paradis E. & Schliep K. (2019) ape 5.0: an environment for modern phylogenetics and evolutionary analyses in R. *Bioinformatics.* **35**, 526-8.

Pavoine, S. (2020) An r package to analyse biodiversity in ecology. *Methods Ecol. Evol.* **11**(9), 1106-1112.

Pavoine, S., Vallet, J., Dufour, A. B., Gachet, S. & Daniel, H. (2009) On the challenge of treating various types of variables: application for improving the measurement of functional diversity. *Oikos.* **118**(3), 391-402.

Pearman, P. B. (1997) Correlates of amphibian diversity in an altered landscape of Amazonian Ecuador. *Conserv. Biol.* **11**, 1211-1225

Petchey, O. L. & Gaston, K. J. (2002) Functional Diversity (FD), species richness, and community composition. *Ecol. Lett.* **5**, 402-411.

Prado, V. H. & Rossa-Feres, D. C. (2014) The influence of niche and neutral processes on a neotropical anuran metacommunity. *Austral Ecol.* **39**(5), 540-547.

R Development Core Team. 2016. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing ([www.R-project.org](http://www.R-project.org)).

Ribeiro, J. W. Jr., Lima, A. P. & Magnusson, W. E. 2012. The effect of riparian zones on species diversity of frogs in Amazonian forests. *Copeia.* **2012**(3): 375-381.

Ribeiro, L. F., Blackburn, D. C., Stanley, E. L., Pie, M. R.. & Bornschein, M. R. (2017) Two new species of the *Brachycephalus pernix* group (Anura: Brachycephalidae) from the state of Paraná, southern Brazil. *PeerJ.* **5**, e3603.

Ricotta, C. (2005) Through the jungle of biological diversity. *Acta Biotheor.* **53(1)**, 29-38.

Rodríguez., L. & Duellman, W. (1994) Guide to the frogs of the Iquitos Region, Amazonian Peru. Lawrence, Kansas, 103p.

Rojas-Ahumada, D. P., Landeiro, V. L. & Menin, M. (2012) Role of environmental and spatial processes in structuring anuran communities across a tropical rain forest. *Austral Ecol.* **37(8)**, 865-873.

Schalk, C. M., Montana, C. G., Klemish, J. L. & Wild, E. R. (2014) On the diet of the frogs of the Ceratophryidae: synopsis and new contributions. *S. Am. J. Herpetol.* **9(2)**, 90–105.

Schiesari, L., Zuanon, J., Azevedo-Ramos, C., Garcia, M., Gordo, M. Messias, M. & Vieira, E. M. (2003) Macrophyte rafts as dispersal vectors for fishes and amphibians in the Lower Solimões River, Central Amazon. *J. Trop. Ecol.* **19(3)**, 333-336.

Shangpliang, P. W., Nongkynrih, S. J. & Hooroo, R. N. K. (2018) Breeding site selection of some anurans in Meghalaya in relation to the physicochemical variables of water. *Int. Multidiscip. Res.* **4(6)**, 16-28.

Silva R. A., Martins I. A. & Rossa-Feres D. C. (2011) Environmental heterogeneity: anuran diversity in homogeneous environments. *Zoologia.* **28**, 610–618.

Smith A. M. & Green M. D. (2005) Dispersal and the metapopulation paradigm in amphibian ecology and conservation: are all amphibian populations metapopulations? *Ecography.* **28(1)**, 110-128.

Starr, C., Taggart, R., Evers, C. & Starr, L. (2015) *Biology: The unity and diversity of life.* Cengage Learning. subtropical Australia. *Restor.* **14**, 67-76.

Sun, Z., Zhao, C., Zhu, W., Zhu, W., Feng, J., Su, S. & Zhao, T. (2021) Microhabitat features determine the tadpole diversity in mountainous streams. *Ecol. Indic.* **126**, 107647.

Tavares-Junior, C., Eskinazi-Sant'Anna, E. M. & Pires, M. R. S. (2020) Environmental drivers of tadpole community structure in temporary and permanent ponds. *Limnologica.* **81**, 125764.

Telles, D. O. C. (2014) Influência dos fatores bióticos e abióticos na estrutura de comunidade de girinos em banco de macrófitas no lago Catalão, Amazônia Central. Dissertação de mestrado, Universidade Federal do Amazonas, Manaus, Amazonas, 46p.

Thabah, C. M., Devi, L. M., Hooroo, R. N. K. & Dey, S. (2018) Morphological alterations in the external gills of some tadpoles in response to pH. *J. Morphol. Sci.* **35(02)**, 142-152.

Touchon, J. C. & Worley, J. L. (2015) Oviposition site choice under conflicting risks demonstrates that aquatic predators drive terrestrial egg-laying. *Proc. Royal Soc.* **282(1808)**, 20150376. <http://dx.doi.org/10.1098/rspb.2015.0376>

Upton, K. (2015) Amphibian diversity in Amazonian flooded forests of Peru. PhD Thesis, University of Kent, Kent.

Upton, K., Warren-Thomas, E., Rogers, I. & Docherty, E. (2014) Amphibian diversity on floating meadows in flooded forests of the Peruvian Amazon. *Herpetol. Rev.* **45(2)**, 209-212.

Vasconcelos, T. S., da Silva, F. R., dos Santos, T. G., Prado, V. H. & Provete, D. B. (2019) Biogeographic Patterns of South American Anurans. 1 ed. Springer Nature Switzerland AG, Switzerland, 149p.

Verberk, W. C., Siepel, H. & Esselink, H. (2008). Life-history strategies in freshwater macroinvertebrates. *Freshw. Biol.* **53(9)**, 1722-1738.

Vignoli, L., Bologna, M. & D'Amen, M. (2007) The effects of temperature and pH on the embryonic development of two species of *Triturus* (Caudata: Salamandridae). *Amphibia-Reptilia*. **28(2)**, 295-300.

Villamarín, F., Moraes, L. J. C. L., Ferrara, C. R., Kaefer, I. L., Werneck, F. P. & de Fraga, R. (in press) Ecology and Conservation of Wetland Amphibians and Reptiles, Reference module in Earth Systems and Environmental Sciences. Elsevier Sci. <https://doi.org/10.1016/B978-0-12-819166-8.00052-9>.

Villéger, S., Mason, N. W. & Mouillot, D. (2008) New multidimensional functional diversity indices for a multifaceted framework in functional ecology. *Ecology*. **89(8)**, 2290-2301.

Vitt, L. J. & Caldwell J. P. (1994) Resource utilization and guild structure of small vertebrates in the Amazon Forest leaf litter. *J. Zool.* **234**, 463-76.

Wake, D. B. & Koo, M. S. (2018) Primer: amphibians. *Current Biology*. **28**: 1221-1242.

Webb, C. O., Ackerly, D. D. & Kembel, S. W. (2008) Phylocom: software for the analysis of community phylogenetic structure and trait evolution. Version 4.0.1. <http://www.phylodiversity.net/phylocom/>. *community ecology. Annual Review of Ecology and Systematics*. 33, 475–505.

Weiher, E. & Keddy, P. A. (1995) Assembly rules, null models, and trait dispersion: new questions from old patterns. *Oikos*. **74(1)**, 159-164.

Whitfeld, T. J. S., Kress, W. J., Erickson, D. L. & Weiblen, G. D. (2012) Change in community phylogenetic structure during tropical forest succession: evidence from New Guinea. *Ecography*. **35**, 821–830.

Zuchi, N., Röpke, C., Shibuya, A., Farago, T., Carmona, M., Zuanon, J. & Amadio, S. (2020) Length-weight relationship of fish species from Central Amazon floodplain. *J. Appl. Ichthyol.* **36(6)**, 837-841.

Zuur, A., Ieno, E. N., Walker, N., Saveliev, A. A. & Smith, G. M. (2009) *Mixedeffects models and extensions in ecology with R*. New York. NY, Springer, 574p.